

Global Epidemiology and Antifungal Resistance Trends of Candida Species (2000-2025): A Systematic Review of Species Distribution, Outbreaks, and Emerging Multidrug Resistance

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Abstract

Background: The objective is to synthesize data on the distribution, antifungal resistance trends of *Candida* spp., hospital outbreaks and emergence of multidrug resistance across the world from 2000 to 2025. **Material and Methods:** A systematic review was carried out following PRISMA 2020 guidelines. Systematic reviews, meta-analyses, multi-country surveillance reports, cohort studies, cross-sectional studies and outbreak investigations were identified via PubMed/MEDLINE, Embase, Web of Science and Scopus databases for *Candida* species-level identification from human clinical infections. Clinical specimen source, geographic distribution, antifungal susceptibility, mechanisms of antifungal resistance, outbreak characteristics and outcomes of death were retrieved. Due to the differences in the study platforms, breakpoints, clinical settings and surveillance techniques across regions, findings were summarized narratively and supplemented with pooled estimates from eligible published meta-analyses. **Results:** After duplicates were removed, 139 studies contributed to the qualitative synthesis from 2864 records of which 755 full-text articles were evaluated. *Candida albicans* remained important, but non-*albicans* *Candida* species collectively exceeded *Candida albicans* in several regions and clinical settings. *Candida tropicalis*, *Candida parapsilosis*, *Candida glabrata*, and *Candida auris* showed the most clinically relevant shifts. Fluconazole resistance was particularly high in *Candida auris* and increased in *Candida tropicalis* in parts of Asia. Echinocandin resistance was increasingly reported in *Candida glabrata* and *Candida parapsilosis*. *Candida auris* caused multiple healthcare-associated outbreaks because of environmental persistence, delayed identification, multidrug resistance, and inter-facility transmission. **Conclusion:** *Candida* epidemiology has changed substantially over the last 25 years. Species-level identification, standardized susceptibility testing, active surveillance, infection prevention, and antifungal stewardship are essential to reduce morbidity, mortality, and outbreak risk from resistant *Candida* infections.

Keywords: *Candida*; candidemia; antifungal resistance; *Candida auris*; systematic review.

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INTRODUCTION

Candida species are among the most important fungal pathogens in clinical medicine, producing disease that ranges from superficial mucocutaneous infection to life-threatening invasive candidiasis.^[1] Historically, *Candida albicans* was the dominant etiological agent; however, surveillance from the last two decades demonstrates a gradual shift toward non-*albicans* *Candida* species, particularly *C. tropicalis*, *C. parapsilosis*, *C. glabrata*, and *C. krusei*.^[2,3] This transition has been linked to broader antifungal exposure, larger immunocompromised populations, increased use of central venous catheters and other invasive devices, and prolonged survival of critically ill patients in intensive care units.^[4,5] The discovery of *Candida auris* in 2009 drastically altered the global mycological landscape.^[6] Some of the most interesting characteristics of *C. auris* are its ability to develop resistance to fluconazole, resistance to echinocandins and amphotericin B, environmental persistence and efficient transmission in healthcare settings.^[7,8] *C. auris* is known as a critical priority

pathogenic fungus and its clinical importance and public health significance indicated by the World Health Organization classification.^[9]

The mortality risk due to invasive candidiasis is still significant, especially for critically ill, immunosuppressed, neonatal, and postoperative patients.^[10] Species susceptibility profile plays a key role in therapeutic decision, as it relies heavily on the accurate species identification. *C. krusei* is intrinsically resistant to fluconazole; *C. glabrata* also tends to have decreased azole susceptibility, plus emerging echinocandin susceptibility, and *C.*

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parapsilosis can have decreased echinocandin susceptibility, as a result of FKS polymorphisms.^[11-13] Differences between Clinical and Laboratory Standards Institute and European Committee on Antimicrobial Susceptibility Testing breakpoints, as well as variation in laboratory capacity, complicate cross-study comparisons.^[14]

Reliable global estimation is also limited by underreporting from regions with restricted diagnostic access and weak mycology surveillance infrastructure.^[15] An updated synthesis is therefore needed to integrate evidence from systematic reviews, surveillance networks, cohort studies, and outbreak reports. This systematic review summarizes global and regional species distribution, antifungal resistance trends, hospital outbreaks, and mortality patterns of clinically relevant *Candida* species from 2000 to 2025.

MATERIALS AND METHODS

Protocol and reporting standards

This systematic review was prepared in accordance with the PRISMA 2020 statement.^[16] A protocol defining the review question, eligibility criteria, search strategy, data extraction plan, and quality assessment approach was prepared before screening. The protocol was not registered.

Eligibility criteria

Eligible studies reported species-level identification of *Candida* isolates obtained from human clinical infections. Bloodstream, urinary, oral, esophageal, vulvovaginal, cutaneous, wound, device-associated, respiratory, central nervous system, and deep-seated infections were considered. Eligible designs included systematic reviews, meta-analyses, multicentric surveillance reports, large cohort studies, cross-sectional prevalence studies, and outbreak investigations. Single case reports were excluded except when they described rare species, early *Candida auris* detections, or sentinel events with clear epidemiological importance. Purely in vitro studies without clinical data were excluded.

Information sources and search strategy

PubMed/MEDLINE, Embase, Web of Science, and Scopus were searched for articles published between 1 January 2000 and 7 November 2025. Search terms included combinations of “*Candida*,” “candidemia,” “invasive candidiasis,” “*Candida auris*,” “non-*albicans Candida*,” “antifungal resistance,” “azole resistance,” “echinocandin resistance,” “systematic review,” “meta-analysis,” “surveillance,” and “outbreak.” Reference lists of eligible articles and major surveillance reports were screened to identify additional relevant records.^[17-19]

Study selection and data extraction

Two reviewers independently screened titles and abstracts. Full texts were reviewed when records appeared relevant or when eligibility was uncertain. Disagreements were resolved by discussion and, when needed, by consultation with a third reviewer [20]. Extracted data included study year, country or region, study design, patient group, specimen type, identification method, *Candida* species distribution, antifungal susceptibility method, breakpoints used, resistance proportions, resistance mechanisms, outbreak setting, infection-control findings, and mortality outcomes.

Quality assessment

The quality of systematic reviews was evaluated using the AMSTAR 2.^[21] Appraisal of observational studies was conducted on relevant domains of the Newcastle-Ottawa Scale and Joanna Briggs Institute checklists such as representativeness, sampling clarity, laboratory confirmation, ascertaining of outcome and completeness of reporting.^[22,23] Case definition, species confirmation, environmental investigation, typing method, and description of control measures were evaluated in the outbreak reports.

Data synthesis

Due to the differences in geographical location, clinical setting, methods of diagnosis, susceptibility breakpoints, and reporting format of the included studies, no de novo meta-analysis was conducted. Rather, the results of the eligible published meta-analyses were collated and large surveillance and multicenter studies.^[24-26] were included. Data has been sorted by clinical specimen type and species, by antifungal class, by resistance mechanism, by outbreak setting and by reported mortality.

Ethical considerations

Approval by an ethics committee was not warranted as this review was based on aggregate data that do not involve any contact with patients, patient data or animal experiments.

RESULTS

Study selection: There were 3215 records found in the database search. Of the 2864 records screened by title and abstract after duplicate removal, 755 full-text articles were retrieved and assessed for eligibility. Of these, 616 were excluded for not meeting the eligibility criteria. Lastly, 139 studies provided data for the qualitative synthesis – systematic reviews, meta-analyses, surveillance reports, cohort studies and outbreak investigations. The PRISMA flow diagram is shown in Figure 1.

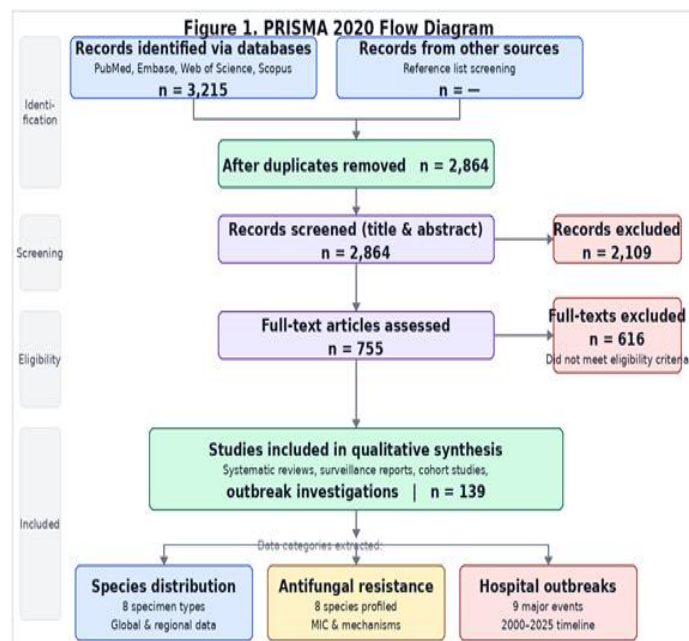


Figure 1: PRISMA 2020 flow diagram showing study selection for the systematic review of *Candida* epidemiology and antifungal resistance trends from 2000 to 2025.

Species distribution across clinical specimens

Species distribution varied by specimen type, patient population, and region. Bloodstream infections continued to be dominated by *C. albicans*, *C. tropicalis*, *C. parapsilosis*, and *C. glabrata*, with *C. auris* becoming increasingly important after 2010 in several countries.^[27-30] A consistent shift toward non-*albicans* *Candida* species was evident, especially in Asia and South America.^[30,31]

Urinary isolates were commonly represented by *C. tropicalis* and *C. albicans*, with *C. tropicalis* more frequently reported among patients with diabetes mellitus, catheterization, and prolonged hospitalization.^[32] Vulvovaginal candidiasis remained predominantly associated with *C. albicans*, although *C. glabrata* was increasingly recognized in recurrent

disease and after azole exposure.^[33,34] Oral isolates consisted mainly of *C. albicans*, *C. glabrata*, *C. tropicalis*, and *C. dubliniensis*, with *C. dubliniensis* more frequently reported in immunosuppressed and human immunodeficiency virus-infected populations.^[35]

Colonization was mostly associated with respiratory isolation, despite reports of invasive diseases among the most ill ventilated patients.^[36] *C. tropicalis* and *C. parapsilosis* occurred frequently in burn wounds, postoperative tissue infections and device related infections.^[37] Non-*albicans* *Candida* species like *C. parapsilosis* and *C. guilliermondii* contributed more to the problem of cutaneous and nail infections.^[38] Central nervous system infections were rare but very serious, primarily in neonates and neurosurgical patients.^[39]

Table 1: Distribution of major Candida species by clinical specimen type, 2000-2025

Sample type	Predominant species	Major epidemiological trend	Key references
Bloodstream infections	<i>C. albicans</i> , <i>C. tropicalis</i> , <i>C. parapsilosis</i> , <i>C. glabrata</i> , <i>C. auris</i>	Declining dominance of <i>C. albicans</i> with increasing non- <i>albicans</i> <i>Candida</i> ; <i>C. auris</i> emerged as a multidrug-resistant bloodstream pathogen after 2010.	[27-30]
Urine	<i>C. tropicalis</i> , <i>C. albicans</i> , <i>C. glabrata</i> , <i>C. krusei</i>	<i>C. tropicalis</i> increased in catheterized, diabetic, and hospitalized patients; azole resistance was increasingly reported.	[32]
Vaginal swabs	<i>C. albicans</i> , <i>C. glabrata</i> , <i>C. krusei</i> , <i>C. parapsilosis</i>	<i>C. albicans</i> remained dominant; <i>C. glabrata</i> was associated with recurrent vulvovaginal candidiasis and prior azole exposure.	[33,34]
Oral cavity	<i>C. albicans</i> , <i>C. glabrata</i> , <i>C. tropicalis</i> , <i>C. dubliniensis</i>	<i>C. dubliniensis</i> was reported more frequently in immunosuppressed populations; <i>C. glabrata</i> and <i>C. tropicalis</i> increased among denture users and older adults.	[35]
Respiratory samples	<i>C. albicans</i> , <i>C. tropicalis</i> , <i>C. glabrata</i>	Most isolates represented colonization; invasive disease was reported mainly in critically ill and immunocompromised patients.	[36]
Wound and tissue	<i>C. albicans</i> , <i>C. tropicalis</i> , <i>C. parapsilosis</i>	<i>C. tropicalis</i> was prominent in burn wound infection; <i>C. parapsilosis</i> was frequent in postsurgical and catheter-associated infections.	[37]
Nail and skin	<i>C. albicans</i> , <i>C. parapsilosis</i> , <i>C. guilliermondii</i>	Non- <i>albicans</i> <i>Candida</i> species gained importance; topical azole exposure may contribute to resistance selection.	[38]
CNS/CSF	<i>C. albicans</i> , <i>C. parapsilosis</i> , <i>C. glabrata</i>	Rare but clinically severe; described mainly in neonates, neurosurgical patients, and immunocompromised hosts.	[39]

Abbreviations: CNS, central nervous system; ICU, intensive care unit; MIC, minimum inhibitory concentration; NAC, non-*albicans* *Candida*; NICU, neonatal intensive care unit.

Antifungal resistance trends

Species and geographical variability was observed in resistance pattern to fungicides. The global surveillance data showed low levels of resistance to fluconazole, echinocandin, and amphotericin B for *C. albicans*.^[40] In contrast, *C. tropicalis* demonstrated an appreciable rise in fluconazole resistance in Asia, which was due to ERG11 mutations, efflux pump activity, biofilm formation, and antifungal selection pressure.^[31,41]

A reduction in susceptibility to azoles and selection of echinocandin resistance due to FKS1 and FKS2 mutations

were observed in *C. glabrata*.^[42,43] *C. parapsilosis* was a growing concern due to decreasing echinocandin susceptibility and resistance signals reported in Europe and Latin America.^[44] *C. auris* had the most alarming profile, with high fluconazole resistance; variable echinocandin resistance, and amphotericin B resistance dependent upon the clade.^[28,29,45]

These patterns facilitate simple, routine species identification and susceptibility testing of clinically relevant isolates, with special focus on those from bloodstream infection, neonatal units, transplant services and intensive care units.^[47]

Table 2: Antifungal resistance trends among major Candida species, 2000-2025

Species	Fluconazole resistance	Echinocandin resistance	Amphotericin B resistance	Major mechanisms and clinical implications	References
<i>C. albicans</i>	<10%; generally stable	<2%	<1%	ERG11 alteration and efflux pump activation in resistant isolates; resistance remains relatively uncommon in most surveillance datasets.	[40]
<i>C. tropicalis</i>	20-45% in several Asian reports; rising trend	<5%	<2%	ERG11 mutations, efflux pump activity, and biofilm-mediated tolerance; important in candidemia and candiduria.	[31,41]
<i>C. glabrata</i>	15-30%; reduced susceptibility and multidrug resistance	5-10%; mainly linked to FKS1/FKS2	<1%	Intrinsic azole tolerance with acquired resistance; echinocandin resistance threatens first-line therapy.	[42,43]

	increasingly reported	mutations			
C. parapsilosis	10-20%	5-12%; increasing in selected regions	<1%	FKS1 polymorphisms and biofilm formation on plastic devices; important in catheter-related and neonatal infection.	[44]
C. krusei	Intrinsic fluconazole resistance	<2%	1-4%	Altered azole target enzyme; species-level identification is essential before azole therapy.	[11,12]
C. auris	70-90%; frequently pan-azole resistant	5-10%; increasing in some clades and outbreaks	5-35%	ERG11 mutations, ERG pathway alterations, ABC transporter upregulation, environmental persistence, and clonal spread.	[28,29,45]
C. dubliniensis	<5%; uncommon	Rare	Rare	Closely related to C. albicans; resistance remains sporadic but should be monitored in recurrent infection.	[46]
C. guilliermondii	10-20%	2-5%	<2%	Reduced azole susceptibility and altered FKS gene signals reported in selected datasets.	[47]

Abbreviations: CNS, central nervous system; ICU, intensive care unit; MIC, minimum inhibitory concentration; NAC, non-albicans Candida; NICU, neonatal intensive care unit.

Hospital outbreaks

Infection outbreaks with Candida species were mainly associated with heavy device use, environmental contamination and susceptible hosts. Initial reports often embraced C. parapsilosis in neonatal and adult intensive care units due to hand transmission and catheter biofilms which persist.^[48] In India, clusters of C. tropicalis have been reported in surgical and medical intensive care units with molecular evidence of clonal spread, as well as high level of

fluconazole resistance.^[49]

Since then, C. auris was the most problematic outbreak organism with large HA outbreaks reported in South Africa, UK, USA, India, Pakistan, Europe and Latin America (50-53). Some organisms have been identified only late, causing colonization to be slow to occur, survival in the environment and resistance to the commonly used antifungals. This allowed dissemination to be rapid between wards and institutions.^[62-66]

Table 3: Major hospital outbreak patterns due to Candida species, 2000-2025

Period	Country/region	Species	Setting/source	Key outbreak findings	References
2000-2006	United States and Europe	C. parapsilosis	NICU/ICU; catheter-related infection	Hand transmission and catheter biofilms sustained outbreaks among neonates and critically ill patients.	[48]
2009-2013	India	C. tropicalis	Surgical and medical ICUs; urinary catheters	Fluconazole resistance and clonal spread reported in high-risk units.	[31,49]
2011-2016	South Africa	C. auris	ICUs and bloodstream infections	Large-scale healthcare-associated emergence with high azole resistance and significant mortality.	[50,63]
2012-2017	United Kingdom	C. auris	Critical care units	Environmental contamination, surface persistence, and fomite transmission documented.	[51,62]
2013-2019	United States	C. auris	Long-term care and acute-care hospitals	Multistate transmission with emerging echinocandin resistance signals.	[52,64]
2014-2020	Spain and Italy	C. parapsilosis	Surgical wards and catheter use	Echinocandin-resistant isolates and hand hygiene gaps reported.	[60,61]
2015-2023	India and Pakistan	C. auris	ICUs and ventilator-associated settings	Clade I dominance, hospital-to-hospital dissemination, and high mortality were reported.	[28,45,66]
2017-2022	Latin America	C. parapsilosis and C. glabrata	NICUs and surgical units	Persistent colonization and environmental reservoirs contributed to recurrent clusters.	[19]
2018-2025	Global	C. auris	Multicountry healthcare facilities	Global recognition as a critical-priority fungal pathogen with continued outbreaks despite control measures.	[9,53,77]

Abbreviations: CNS, central nervous system; ICU, intensive care unit; MIC, minimum inhibitory concentration; NAC, non-albicans Candida; NICU, neonatal intensive care unit.

DISCUSSION

There is an evident shift in the epidemiology of Candida over the last two and a half decades as shown in this systematic review. C. albicans is still a significant pathogen, but there are now several datasets that indicate that non-albicans Candida species are now contributing a greater proportion, particularly C. tropicalis, C. parapsilosis, C. glabrata and C. auris.^[54,55] Clinically significant because non-albicans Candida may be less susceptible to azoles, echinocandins or both.

The emergence of C. tropicalis in Asia is of particular importance in India and its neighbouring nations. It has been

consistently reported to be associated with diabetes mellitus, catheterization, intensive care exposure and urinary or blood stream infection.^[56] The high resistance rates reported in some regions, which are higher than 40%, indicate a potential for clonal expansion, ERG11-mediated resistance, biofilm tolerance, and extensive azole use creating the appearance of a high level of resistance. The results confirm local antibiogram-based antifungal policies as opposed to using global averages. In many high-income environments, C. glabrata is a significant pathogen which yields a low susceptibility to azoles and ability to acquire echinocandin resistance. Clinically, the FKS mutations are important as the echinocandins are used as first-line treatment

for invasive candidiasis [58,59]. Likewise, *C. parapsilosis* has been considered a strong correlate to catheter related infection and neonatal sepsis. In Europe and Latin America, reports of *C. parapsilosis* isolates resistant to echinocandin drugs were particularly noteworthy because *C. parapsilosis* is a species that is known to be capable of forming biofilms on plastic surfaces and spread via hand-to-hand contact.^[60,61] *C. auris* represents the most significant healthcare-associated threat among *Candida* species in recent decades. It is unique among *Candida* spp. for its ability to survive in the environment, to colonize without causing clinical symptoms, to be multi-drug resistant, and to be transmitted in healthcare settings.^[62-66] Therapeutic uncertainty exists due to the high level of fluconazole resistance, and the relative lack of echinocandin resistance and the presence of clade-dependent amphotericin B resistance.^[66] The features are the reasons why species identification, patient isolation, contact screening, environmental cleaning (with effective disinfectants) and molecular surveillance are all necessary in outbreak settings.^[53,74-77]

Patterns also need to be considered which are specific to the type of specimen. Although *C. albicans* remains a prominent cause of oral candidiasis, other species of *Candida*, such as *C. dubliniensis*, *C. glabrata* and *C. tropicalis* are gaining significance in immunocompromised patients and individuals wearing dentures.^[35,67] *C. albicans* is still the most common organism involved in RVC, with the other most common species occurring after exposure to azole antifungals, such as *C. glabrata*.^[33,69] Increased Nail and Skin infection recovery of non-*albicans* *Candida* may be due to topical azole utilization and shifting environmental reservoirs.^[70]

The review points to some critical surveillance gaps worldwide. Routine MALDI-TOF mass spectrometry, molecular identification and standardized susceptibility testing is not available in many LMIC areas. The delayed response of infection control measures may be caused by the confusion of the uncommon *Candida* species, *C. auris* and others, with other species.^[75] Thus, investment in lab capacity, uniform break points for the AGPs, antifungal stewardship, and genomic epidemiology is necessary to identify resistant clones early and prevent spread between facilities.^[76,77]

Strengths and limitations

This review's major limitation is that the evidence was synthesized across specimen types and regions, as well as across antifungal classes and outbreak situations, which likely introduces heterogeneity into the evidence. It combines with systematic reviews, large surveillance studies and outbreak investigations to give an overview of the changing epidemiology of *Candida* in a clinically useful way. Main limitations include: study design, laboratory methodology, susceptibility breakpoints and reporting quality. Certain areas are under-represented, and a number of resistance estimates were based on monitored networks rather than population-based networks. A de novo meta-analysis was not performed because of this heterogeneity.

CONCLUSION

There have been significant changes in *Candida* epidemiology between 2000 and 2025. However, non-*albicans* *Candida* species are becoming more common, and resistance to antifungals is growing in clinically relevant species like *C. tropicalis*, *C. glabrata*, *C. parapsilosis* and *C. auris*. Multidrug resistance, environmental stability and outbreak potential make *C. auris* a significant threat to healthcare. Specifics of species identification, standardized susceptibility testing, targeted antifungal stewardship and strong infection-prevention strategies form the basis of good outcomes. Increased surveillance and genomic epidemiology is required to track emergence of resistance and decrease the burden of invasive candidiasis worldwide.

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Conflicts of interest

There are no conflicts of interest.

REFERENCES

1. Calderone RA, Clancy CJ. *Candida and Candidiasis*. 2nd ed. Washington, DC: ASM Press; 2012.
2. Guinea J. Global trends in the distribution of *Candida* species causing candidemia. *Clin Microbiol Infect*. 2014;20 Suppl 6:5-10.
3. Lamoth F, Lockhart SR, Berkow EL, Calandra T. Changes in the epidemiology of invasive candidiasis. *J Antimicrob Chemother*. 2018;73:i4-i13.
4. Pfaller MA, Diekema DJ. Epidemiology of invasive candidiasis: a persistent public health problem. *Clin Microbiol Rev*. 2007;20:133-163.
5. Kullberg BJ, Arendrup MC. Invasive candidiasis. *N Engl J Med*. 2015;373:1445-1456.
6. Satoh K, Makimura K, Hasumi Y, et al. *Candida auris* sp. nov., a novel ascomycetous yeast. *Microbiol Immunol*. 2009;53:41-44.
7. Chowdhary A, Sharma C, Meis JF. *Candida auris*: a rapidly emerging cause of hospital-acquired multidrug-resistant fungal infections. *J Infect*. 2017;75:1-11.
8. Lockhart SR, Etienne KA, Vallabhaneni S, et al. Simultaneous emergence of multidrug-resistant *Candida auris* on three continents. *Clin Infect Dis*. 2017;64:134-140.
9. World Health Organization. WHO fungal priority pathogens list to guide research, development and public health action. Geneva: World Health Organization; 2022.
10. Pappas PG, Lionakis MS, Arendrup MC, Ostrosky-Zeichner L, Kullberg BJ. Invasive candidiasis. *Nat Rev Dis Primers*. 2018;4:18026.
11. Arendrup MC. Update on antifungal resistance in *Candida* species. *Curr Opin Infect Dis*. 2013;26:493-500.
12. Perlin DS. Mechanisms of echinocandin antifungal drug resistance. *Ann N Y Acad Sci*. 2015;1354:1-11.
13. Pfaller MA, Diekema DJ. Progress in antifungal susceptibility testing. *Clin Microbiol Rev*. 2020;33:e00069-19.
14. Arendrup MC, Meletiadis J, Mouton JW, et al. EUCAST and CLSI breakpoints for *Candida*. *Clin Microbiol Infect*. 2017;23:676-683.
15. Bongomin F, Gago S, Oladele RO, Denning DW. Global and multinational prevalence of fungal diseases. *Eur J Clin Microbiol Infect Dis*. 2017;36:1051-1057.
16. Page MJ, McKenzie JE, Bossuyt PM, et al. The PRISMA 2020 statement. *BMJ*. 2021;372:n71.
17. Pfaller MA, Diekema DJ. Global surveillance of *Candida* infections.

- J Clin Microbiol. 2010;48:437-441.
18. Turner SA, Butler G. The Candida pathogenic species complex. *PLoS Pathog.* 2014;10:e1004366.
 19. Colombo AL, Guinea J, de Almeida Junior JN, et al. Epidemiology of candidemia. *Clin Microbiol Infect.* 2017;23:673-680.
 20. Higgins JPT, Green S, editors. *Cochrane Handbook for Systematic Reviews of Interventions.* London: The Cochrane Collaboration; 2011.
 21. Shea BJ, Reeves BC, Wells G, et al. AMSTAR 2: a critical appraisal tool for systematic reviews. *BMJ.* 2017;358:j4008.
 22. Wells GA, Shea B, O'Connell D, et al. The Newcastle-Ottawa Scale for assessing the quality of nonrandomised studies in meta-analyses. Ottawa: Ottawa Hospital Research Institute; 2013.
 23. Joanna Briggs Institute. JBI critical appraisal tools. Adelaide: Joanna Briggs Institute; 2020.
 24. Benedict K, Jackson BR, Chiller T, Beer KD. Estimation of fungal disease burden. *Open Forum Infect Dis.* 2019;6(Suppl 4):S14-S21.
 25. Lamoth F. Antifungal susceptibility testing developments. *Antimicrob Agents Chemother.* 2019;63:e01733-18.
 26. Silva S, Negri M, Henriques M, Oliveira R, Williams DW, Azeredo J. Candida biofilms: a review. *J Dent.* 2011;39:879-887.
 27. Pfaller MA, Diekema DJ. Role of sentinel surveillance. *J Clin Microbiol.* 2007;45:880-885.
 28. Chowdhary A, Prakash A, Sharma C, et al. A multicentre study of Candida auris in India. *Mycoses.* 2018;61:314-317.
 29. Lockhart SR, Berkow EL, Chow N, Welsh RM. Candida auris for clinicians. *J Clin Microbiol.* 2022;60:e00380-22.
 30. Arendrup MC, Dzajic E, Jensen RH, et al. Epidemiology of candidemia in Asia. *Clin Microbiol Infect.* 2013;19:946-953.
 31. Kothavade RJ, Kura MM, Valand AG, Panthaki MH. Epidemiology of Candida tropicalis in India. *Mycoses.* 2010;53:60-69.
 32. Achkar JM, Fries BC. Candiduria review. *Clin Microbiol Rev.* 2010;23:253-273.
 33. Sobel JD. Recurrent vulvovaginal candidiasis. *Lancet.* 2016;4:e262-e272.
 34. Deorukhkar SC, Saini S, Mathew S. Non-albicans Candida in vulvovaginal candidiasis. *J Obstet Gynaecol India.* 2017;67:226-229.
 35. Samaranyake LP, Keung Leung W, Jin L. Oral candidiasis. *J Oral Biosci.* 2009;51:2-7.
 36. Meersseman W, Lagrou K, Maertens J, et al. Invasive candidiasis in the intensive care unit. *Clin Infect Dis.* 2009;48:503-511.
 37. Eggimann P, Garbino J, Pittet D. Candida burn wound infections. *Lancet Infect Dis.* 2015;15:611-622.
 38. Gupta AK, Versteeg SG, Shear NH. Onychomycosis updates. *J Am Acad Dermatol.* 2017;76:935-949.
 39. Bongomin F, Oladele RO, Gago S, Moore CB, Richardson MD. Central nervous system Candida infections. *J Fungi.* 2021;7:431.
 40. Pfaller MA, Castanheira M, Messer SA, Jones RN. Global antifungal surveillance. *Antimicrob Agents Chemother.* 2019;63:e00840-19.
 41. Singh A, Chakrabarti A, Bhattacharya S, et al. ERG11 mutations in Candida tropicalis. *J Med Microbiol.* 2022;71:001553.
 42. Wiederhold NP. Antifungal resistance in Candida glabrata. *Clin Infect Dis.* 2017;64:1-6.
 43. Pfaller MA, Diekema DJ. FKS mutations and echinocandin resistance. *Clin Microbiol Rev.* 2020;33:e00103-19.
 44. Grossman NT, Chiller TM, Lockhart SR. Candida parapsilosis resistance. *Open Forum Infect Dis.* 2015;2:ofv163.
 45. Lockhart SR, Welsh RM, Lewis J. Candida auris resistance evolution. *J Antimicrob Chemother.* 2023;78:5-14.
 46. Sullivan DJ, Moran GP, Pinjon E. Candida dubliniensis. *Rev Iberoam Micol.* 2016;33:66-74.
 47. Espinel-Ingroff A. Global trends in Candida resistance. *Med Mycol.* 2020;58:106-117.
 48. Trofa D, Gacser A, Nosanchuk JD. Candida parapsilosis review. *Clin Microbiol Rev.* 2008;21:606-625.
 49. Sharma C, Kumar N, Pandey R, Meis JF, Chowdhary A. Indian Candida tropicalis outbreaks. *Med Mycol.* 2013;51:557-562.
 50. Magobo RE, Corcoran C, Seetharam S, Govender NP. First South African Candida auris outbreak. *Mycoses.* 2014;57:257-265.
 51. Schelenz S, Hagen F, Rhodes JL, et al. First hospital outbreak of Candida auris in the United Kingdom. *Clin Infect Dis.* 2016;62:1214-1220.
 52. Vallabhaneni S, Kallen A, Tsay S, et al. Investigation of the first seven reported cases of Candida auris, a globally emerging invasive, multidrug-resistant fungus, United States, May 2013-August 2016. *Clin Infect Dis.* 2016;63:95-102.
 53. World Health Organization. WHO fungal priority pathogens list to guide research, development and public health action. Geneva: World Health Organization; 2022. Available from: <https://www.who.int/publications/i/item/9789240060241>
 54. Pfaller MA, Castanheira M. Global emergence of non-albicans Candida species. *Clin Microbiol Rev.* 2020;33:e00134-19.
 55. Guinea J, Puig-Asensio M, et al. Epidemiology of candidemia: a shift to non-albicans Candida. *Clin Microbiol Infect.* 2014;20 Suppl 6:5-10.
 56. Tan TY, Hsu LY, et al. Candida tropicalis in Asia. *J Med Microbiol.* 2016;65:256-264.
 57. Singh A, Singh PK, et al. ERG11-mediated azole resistance. *J Med Microbiol.* 2022;71:001553.
 58. Healey KR, Perlin DS. FKS mutations in Candida glabrata. *Antimicrob Agents Chemother.* 2018;62:e02233-17.
 59. Alexander BD, Johnson MD. Rising echinocandin resistance. *Curr Opin Infect Dis.* 2020;33:520-528.
 60. Garcia-Hermoso D, Canton R, et al. Echinocandin-resistant Candida parapsilosis. *Clin Microbiol Infect.* 2018;24:1109-1110.
 61. Marcos-Zambrano LJ, Escribano P, et al. FKS polymorphisms in Candida parapsilosis. *J Antimicrob Chemother.* 2021;76:1234-1242.
 62. Jeffery-Smith A, Taori SK, et al. Candida auris outbreaks in the United Kingdom. *Clin Infect Dis.* 2018;66:1244-1252.
 63. Govender NP, Magobo RE, et al. South African Candida auris epidemic. *Mycoses.* 2016;59:1-7.
 64. Tsay S, Welsh RM, et al. United States Candida auris surveillance. *MMWR.* 2020;69:1-6.
 65. Osei Sekyere J. Candida auris resistance and outbreaks. *Infect Drug Resist.* 2020;13:1173-1180.
 66. Welsh RM, Sexton DJ, et al. Global antifungal resistance in Candida auris. *J Infect Dis.* 2023;227:1021-1029.
 67. Sullivan DJ, Moran GP. Candida dubliniensis in oral candidiasis. *Med Mycol.* 2011;49:1-16.
 68. Pappas PG, Lionakis MS. Central nervous system candidiasis review. *Nat Rev Dis Primers.* 2018;4:18026.
 69. Goncalves B, Ferreira C, et al. Vaginal Candida species trends. *Microorganisms.* 2021;9:778.
 70. Bitar D, Cassagne C, et al. Non-albicans Candida species in superficial mycoses. *Med Mycol.* 2021;59:402-410.
 71. Arendrup MC, Patterson TF. Azole resistance drivers. *Clin Infect Dis.* 2017;65:113-119.
 72. Perlin DS, Rautemaa-Richardson R, Alastruey-Izquierdo A. Echinocandin resistance. *Curr Opin Infect Dis.* 2017;30:520-529.
 73. Arendrup MC, Meletiadis J. Antifungal stewardship. *J Antimicrob*

- Chemother. 2020;75:3308-3314.
74. Sexton DJ, Welsh RM, et al. Environmental persistence of *Candida auris*. *Clin Infect Dis*. 2021;72:e1144-e1150.
75. Hamprecht A, et al. Misidentification of *Candida auris*. *J Clin Microbiol*. 2019;57:e01787-18.
76. Rhodes J, Fisher MC. Global surveillance and genomics in *Candida auris*. *Nat Rev Microbiol*. 2019;17:327-339.
77. Chow NA, de Groot T, Badali H, et al. Genomic epidemiology and antifungal resistance of *Candida auris*. *Clin Microbiol Rev*. 2023;36:e00014-22.