

Urinary Pathogens and Their Antibiotic Resistance Pattern Among Patients in A Tertiary Care Hospital

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Abstract

Background: Urinary Tract Infections (UTIs) are among the most prevalent infections worldwide, affecting [1] [2]. Various factors increase the risk for UTIs, including sexual activity, vaginal infections, diabetes, hygiene practices, obesity, and genetic predisposition [3]. UTIs are mainly caused by Gram-negative organisms like *Escherichia coli*, *Klebsiella* species, *Proteus* species, and *Pseudomonas* species, and Gram-positive bacteria like *Enterococcus* species, *Staphylococcus aureus*, and CONS (*Staphylococcus saprophyticus*). Among these organisms, *E. coli* is the most common causative organism for community-acquired and hospital-acquired UTIs [5]. **Material and Methods:** The study was conducted in the department of microbiology of Teerthanker Mahaveer Medical College and Research Centre from January 2025 to June 2025 after approval (IRB/134/2025 dated 16 June 2025). A total of 3031 samples were processed. Mid-stream clean catch samples or from the catheter port were collected for culture and antibiotic sensitivity testing. Samples were processed using a calibrated loop and inoculated semi quantitatively on a CLED agar plate as per Kass's recommendations [10]. The identification of bacterial isolates and the antibiotic susceptibility test (AST) were performed manually and using an automated method (VITEK 2). HiCrome agar by HiMedia was used to identify *Candida* isolates. **Results:** In our study, 89 OPD and 308 IPD samples were culture-positive. Females were more frequently affected than males. The most common organisms isolated in OPD and IPD samples were *E. coli* (53.9% and 42.2%, respectively), *Klebsiella pneumonia* (12.4%), and *Enterococcus spp.* (12.4%), and *Candida parasilopsis* (11%). Antibiotics that were more sensitive among Gram-negative organisms were Nitrofurantoin, Aminoglycoside, Carbapenems, and Fosfomycin, whereas among Gram-positive organisms, Nitrofurantoin, Linezolid, Teicoplanin, and Vancomycin were more sensitive. There was a significant association between age and *E. coli* isolation ($\chi^2=18.34$, $df=3$, $p=0.001$), and gender-wise, *E. coli* was found more in females (29.7%) than in males (15.1%), with statistical significance ($\chi^2=16.27$, $df=1$, $p=0.001$). **Conclusion:** The current study aims to detect and determine the hospital-based prevalence of UTI, its causative agents, and their antimicrobial susceptibility patterns. This study will also help to create recommendations for determining effective empirical therapy/ antibiotic policy for UTIs while waiting for culture sensitivity results.

Keywords: UTI, AST, Antimicrobial resistance.

Received: 20 August 2025

Revised: 15 September 2025

Accepted: 01 October 2025

Published: 13 October 2025

INTRODUCTION

Urinary Tract Infections (UTIs) are among the most prevalent infections worldwide. They affect the lower urinary tract (bladder and urethra) and the upper urinary tract (kidney and ureter).^[1,2] Although UTIs can affect any age group, including men and women, clinical studies suggest that the overall prevalence of UTIs is significantly higher in women.

The various factors that increase the risk for UTIs include sexual activity, vaginal infections, diabetes, hygiene practices, obesity, and genetic predisposition.^[3]

Bacteria first colonize and then infect the urinary tract, resulting in symptoms such as dysuria, urinary incontinence, and hematuria.^[4]

The clinical presentation may range from an asymptomatic urinary tract infection to a life-threatening illness like sepsis. UTI infections are mainly caused by Gram-negative organisms like *Escherichia coli*, *Klebsiella* species, *Proteus* species, and *Pseudomonas* species, and Gram-positive

bacteria like *Enterococcus* species, *Staphylococcus aureus*, and CONS (*Staphylococcus saprophyticus*). Among these organisms, *E. coli* is the most common causative organism for community-acquired and hospital-acquired UTIs.^[5]

Empirical treatment of UTIs is common in rural areas due to the restricted availability of urine culture testing. This inadequate diagnosis often leads to the misuse of antibiotics. Knowing the area's antibiotic susceptibility pattern is crucial for deciding the empirical selection of antibiotics.^[6,7]

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DOI:
10.21276/amit.2025.v12.i3.119

How to cite this article: Patil P, Rawat S, Farooq U, Sharma SR, Singh S, Vasundhara. Urinary Pathogens and Their Antibiotic Resistance Pattern Among Patients in A Tertiary Care Hospital. Acta Med Int. 2025;12(3):473-478.

Developing nations like India are more vulnerable to the emergence of antimicrobial resistance (AMR) due to poor hygiene, inadequate infection control practices, and a lack of surveillance and implementation of antimicrobial stewardship programs. Self-medication and non-adherence to the prescribed antibiotic regimen also contribute to the emergence of AMR.^[8]

The antibiotic susceptibility pattern for uropathogens varies spatially and temporally in different regions. Thus, antimicrobial susceptibility data are pivotal in generating contemporary and wide-ranging epidemiological data.^[9]

Our study aims to identify the spectrum of uropathogens prevalent in our region and their antimicrobial susceptibility patterns. This study will help shape and reform our institute’s antibiotic policy for better patient outcomes and facilitate a way forward in introducing and implementing antimicrobial stewardship.

MATERIALS AND METHODS

The study was conducted in the department of microbiology of Teerthanker Mahaveer Medical College and Research Centre from January 2025 to June 2025 after approval (IRB/134/2025 dated 16 June 2025). A total of 3031 samples were processed, including those from inpatient (IPD) and outpatient departments (OPD).

Midstream clean catch samples or from the catheter port in case of cauterized patients were collected in a wide-mouth

sterile container for culture and antibiotic sensitivity testing.

Samples were processed using a calibrated loop and inoculated semi quantitatively on a CLED agar plate as per Kass’s recommendations.^[10] The plates were then incubated at 37°C aerobically for 24 hrs. Culture plates were then examined for bacterial growth. A 10⁵CFU/ml bacterial count was taken as a significant count.^[11]

The identification and antibiotic susceptibility test (AST) for the bacterial isolates was done manually and by VITEK 2, an automated system by bioMérieux. HiCrome agar by HiMedia was used to identify Candida isolates.

Antimicrobial agents were tested against the bacterial isolates, and the results were interpreted according to Clinical and Laboratory Standards Institute (CLSI) guidelines 2025.

Data was analyzed using SPSS version 25.0. Categorical variables were expressed as frequency and percentage. Associations between organisms and age groups (df=3) or gender (df=1) were assessed using the Chi-square test; Fisher’s exact test was applied when expected counts were <5. A P value of <0.05 was considered statistically significant.

RESULTS

In our study, 89 OPD and 308 IPD samples were culture positive. Age group-wise, the 20-30 years range shows the maximum infectivity, with 21.3% and 19.2 %, respectively, in OPD and IPD patients. The positivity rate was found to be lowest below 10 years of age and highest above 90 years of age.

Table 1: Distribution of study participants as per age groups

| Age Group (Years) | OPD | | IPD | |
|-------------------|---------------|----------------|---------------|----------------|
| | Frequency (n) | Percentage (%) | Frequency (n) | Percentage (%) |
| 0–9 | 4 | 4.5 | 10 | 3.2 |
| 10–19 | 4 | 4.5 | 16 | 5.2 |
| 20–29 | 19 | 21.3 | 59 | 19.2 |
| 30–39 | 8 | 9.0 | 41 | 13.3 |
| 40–49 | 13 | 14.6 | 48 | 15.6 |
| 50–59 | 16 | 18.0 | 47 | 15.3 |
| 60–69 | 14 | 15.7 | 47 | 15.3 |
| 70–79 | 7 | 7.9 | 30 | 9.7 |
| 80–89 | 3 | 3.4 | 10 | 3.2 |
| 90–99 | 1 | 1.1 | 0 | 0.0 |
| Total | 89 | 100.0 | 308 | 100.0 |
| Mean ±SD | 46.42 | | 20.97 | |

Table 2: Distribution of study participants as per Gender

| Gender | OPD | | IPD | |
|--------|---------------|----------------|---------------|----------------|
| | Frequency (n) | Percentage (%) | Frequency (n) | Percentage (%) |
| Male | 36 | 40.4 | 109 | 35.4 |
| Female | 53 | 59.6 | 199 | 64.6 |
| Total | 89 | 100.0 | 308 | 100.0 |

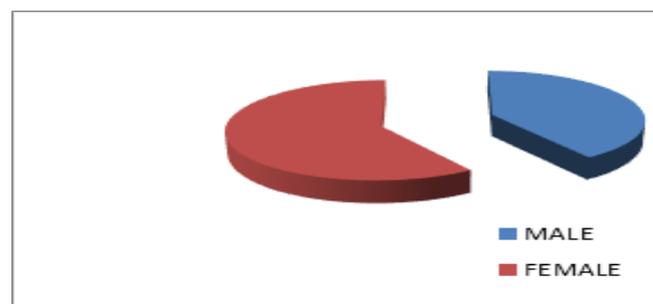


Figure 1: OPD samples

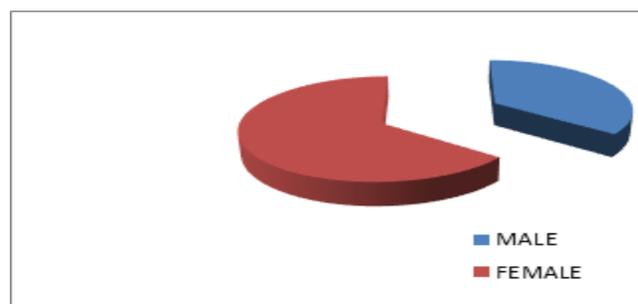


Figure 2: IPD samples

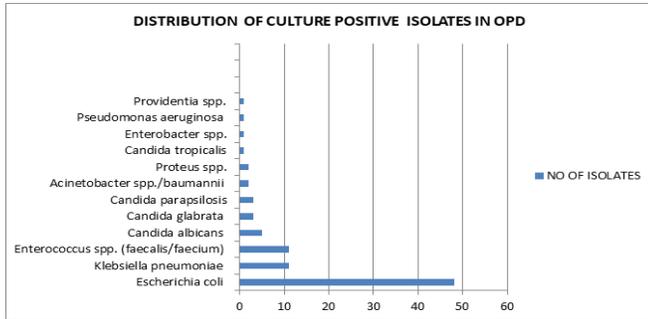


Figure 3: Distribution of Isolated Organisms in OPD

In our study, females were more frequently affected than males. In OPD, 59.6% were females and 40% were males. Whereas in IPD patients, females were 64% positive and males were 35.4%.

In OPD samples, the most common organism isolated was *E.coli* (53.9%), *Klebsiella pneumoniae* (12.4%), and *Enterococcus spp.* (12.4%).

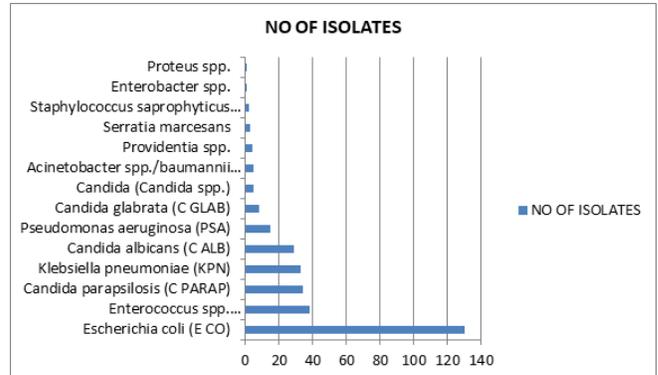


Figure 4: Distribution of Isolated Organisms in IPD

In IPD samples, again *E.coli* (42.2%) was the most predominant isolate followed by *Enterococcus spp.* (12.3%), *Candida parapsilosis* (11%) and *Klebsiella pneumonia* (10.7%).

Table 3: Antibiotic Susceptibility in OPD and IPD patients

| Organism | Most Active (S ≥70%) | High Resistance (R ≥50%) |
|-------------------------------------|---|--|
| <i>Escherichia coli</i> | Nitrofurantoin, Fosfomycin, Amikacin, Chlormaphenicol | Ampicillin, Amoxicillin/Clavulanate, Fluoroquinolones |
| <i>Klebsiella pneumoniae</i> | Amikacin, Imipenem, Meropenem | Ampicillin, Amoxicillin/Clavulanate, Cephalosporins |
| <i>Enterobacter spp.</i> | Amikacin, Gentamicin | Amoxicillin/Clavulanate, Ampicillin/Sulbactam,, Cephalosporins |
| <i>Proteus spp.</i> | Amikacin, Gentamicin, Piperacillin/Tazobactum | Ampicillin, Amoxicillin/Clavulanate |
| <i>Providentia spp.</i> | Amikacin, Imipenem | Ampicillin, Amoxicillin/Clavulanate |
| <i>Serratia marcescens</i> | Amikacin, Gentamicin | Ampicillin, Amoxicillin/Clavulanate |
| <i>Pseudomonas aeruginosa</i> | Amikacin, Imipenem, Meropenem | Ciprofloxacin, Levofloxacin, Ticarcillin/Clavulanate |
| <i>Acinetobacter spp.</i> | Imipenem, Meropenem, Amikacin | Ciprofloxacin, Piperacillin/Tazobactum, Cephalosporins |
| <i>Enterococcus spp.</i> | Vancomycin, Linezolid, Teicoplanin | Ciprofloxacin, Levofloxacin, Erythromycin |
| <i>Staphylococcus saprophyticus</i> | Nitrofurantoin, Linezolid, Vancomycin, Teicoplanin | Ampicillin, Penicillin |

In our study, the antibiotics that are >70 % sensitive among Gram-negative organisms were Nitrofurantoin, Aminoglycoside, Carbapenems, and Fosfomycin, whereas among Gram-positive organisms, Nitrofurantoin, Linezolid, Teicoplanin, and Vancomycin were the most sensitive antibiotics. Furthermore, the most resistant antibiotics were Penicillins, Cephalosporins, and Fluoroquinolones.

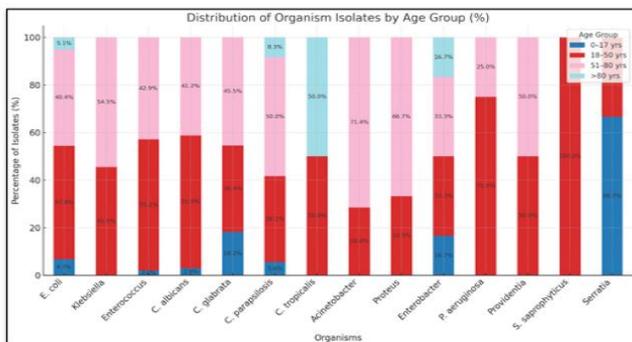


Figure 5: Distribution of isolates age wise analysis

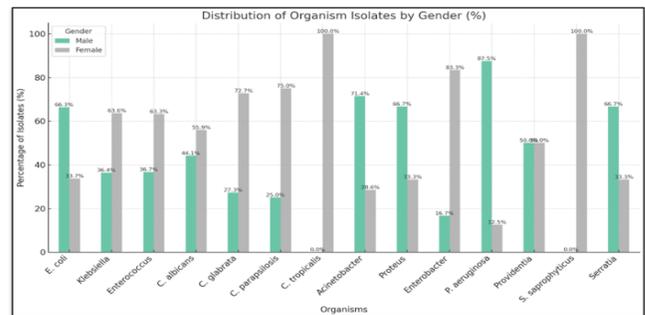


Figure 6: Distribution of isolates gender wise analysis

In the age-wise analysis, *Escherichia coli* was the most common isolate (178; 44.8%), predominantly in the 18–50 years (21.4%) and 51–80 years (18.1%) groups, with a significant association between age and *E. coli* isolation ($\chi^2=18.34$, $df=3$, $p=0.001$). Other organisms include *Klebsiella pneumoniae* (11.1%) and *Enterococcus spp.* (12.3%), *Candida* species were mainly observed in adults, with a few isolates among children and older people.

Table 4: Age wise analysis of *E. coli* isolates

| Organism | 0–17 yrs n (%) | 18–50 yrs n (%) | 51–80 yrs n (%) | >80 yrs n (%) | Total n (%) | Chi-square (χ^2), df | p-value |
|-----------------------------------|----------------|-----------------|-----------------|---------------|-------------|-----------------------------|---------|
| <i>Escherichia coli (E. coli)</i> | 12 (3.0%) | 85 (21.4%) | 72 (18.1%) | 9 (2.3%) | 178 (44.8%) | $\chi^2=18.34$, $df=3$ | 0.001 |

Table 5: Gender wise analysis of *E. coli* isolates

| Organism | Male n (%) | Female n (%) | Total n (%) | Chi-square (χ^2), df | p-value |
|--|-------------|--------------|-------------|-----------------------------|---------|
| <i>Escherichia coli</i> (<i>E. coli</i>) | 118 (29.7%) | 60 (15.1%) | 178 (44.8%) | $\chi^2=16.27$, df=1 | 0.001 |

In the gender-wise comparison, *E. coli* was again predominant (44.8%), occurring more in females (29.7%) than in males (15.1%), with statistical significance ($\chi^2=16.27$, df=1, p=0.001). *Klebsiella pneumoniae*, *Enterococcus spp.*,

and *Candida* species showed relatively higher proportions among females, while *Pseudomonas aeruginosa* was more frequent in males.

Table 6: *E. coli* Antibiotic susceptibility pattern

| Antibiotic | Sensitivity (%) | Antibiotic | Sensitivity (%) |
|-------------------------|-----------------|-------------------------|-----------------|
| Ceftazidime | 11.2 | Netilmicin | 4.4 |
| Cefuroxime | 6 | Tetracycline | 19 |
| Cefotaxime | 7 | Piperacillin–Tazobactam | 36.5 |
| Cefepime | 7.8 | Ticarcillin–Clavulanate | 26.9 |
| Ceftriaxone | 6 | Imipenem | 42 |
| Ampicillin | 6 | Meropenem | 46 |
| Amoxicillin–Clavulanate | 15.7 | Ertapenem | 48 |
| Ampicillin–Sulbactam | 11.2 | Polymyxin B | 100 |
| Norfloxacin | 12 | Colistin | 100 |
| Ofloxacin | 1 | Fosfomycin | 62 |
| Levofloxacin | 8.9 | Nitrofurantoin | 50 |
| Amikacin | 37 | Co-trimoxazole | 29 |
| Tobramycin | 20 | Chloramphenicol | 36 |

Of the 178 *E. coli* isolated, the highest susceptibility was seen in Fosfomycin (62%), followed by Nitrofurantoin (49%), Carbapenems (approximately 45%), Amikacin (37%), Chloramphenicol (36%), and Co-trimoxazole (29%). An overall high level of resistance was observed in fluoroquinolones (approximately 7%) and Cephalosporins (approximately 10%).

DISCUSSION

Urinary tract infections (UTIs) are among the most common infections found worldwide, and the current study highlights the ongoing threat of antimicrobial resistance and the challenges that might be faced in treating such diseases. A multicentric study conducted from 1990 to 2019 has shown a significant increase in the global burden of UTI and mortality rates associated with the rising antimicrobial-resistant organisms.^[12] The prevalence of UTI in the northern region has been seen between 10% to 17%.^[13] The prevalence of UTI in our study was 13% consistent with these findings. However, most studies around Uttar Pradesh have shown much higher prevalence rates.^[14,15]

Uropathogens were higher in number in females (63%) than in males (37%). The distribution was found to be statistically significant in *E. coli* isolates. According to multiple studies, the incidence of UTIs has been higher in females than in males.^[15,16] The anatomy of the female urethra, along with some urological factors such as urinary incontinence, sexual intercourse, and poor personal or toiletry hygiene, may contribute to a higher rate of UTI in females.^[17]

UTI cases were more prevalent in the patients' age group, ranging from 20 to 45 years (47%), which is comparable to some studies.^[16,18] This is also the reproductive age seen in females, highlighting this as an important risk factor for UTIs in young females.

The most common organism isolated in the study was

Escherichia coli (*E. coli*), 54% in OPD patients and 42% in IPD patients. The findings are similar to various studies in and outside India.^[13,15,16,19] Enterobacteriaceae family in UTIs can be linked to their classic morphological factors like pili, fimbriae, adhesins, and p1 blood group receptors, aiding in their colonization and the ability to adhere to the urinary bladder epithelium.^[17] The second most common organism isolated was *Enterococcus* species (n=89; 12.2 % in OPD and n=308; 12.3% in IPD), the finding being consistent with studies from north and south India.^[13,19] Other bacterial isolates obtained in the survey were *Klebsiella pneumoniae*, *Candida* species, *Pseudomonas sp.*, *Acinetobacter sp.*, *Proteus sp.*, *Providentia sp.*, *Enterobacter sp.*, and *Staphylococcus saprophyticus* (CONS). Amongst the isolated organisms, *Candida* species was found to be in higher numbers in IPD patients (OPD- 12%; IPD- 15%), suggesting its pronounced association in hospital-acquired infections or antecedent use of catheters/devices leading to device-associated infections or catheter-associated UTIs (CAUTI). Similar observations have been noted in a study from Uttar Pradesh.^[13] Of the total 178 *E. coli* isolated, the highest susceptibility was seen in Fosfomycin (62%), followed by Nitrofurantoin (50%), Carbapenems (approximately 45%), Amikacin (37%), Chloramphenicol (36%), and Co-trimoxazole (29%). These antibiotics could be more beneficial in treating UTIs in the current setting as the first line of management, except for restricting the use of Carbapenems. However, they are observed to be moderately sensitive (~ 45%), as they are classified under the reserved group category in WHO's AWaRe classification of antimicrobials. Nitrofurantoin susceptibility is comparable to various studies.^[20–22] Many previous studies from India have shown that Nitrofurantoin and Fosfomycin are good initial treatment options.^[17,23,24] Higher susceptibility to Nitrofurantoin may be attributable to its restricted use for acute cystitis, narrow tissue distribution and spectrum of action, small serum concentration, and bactericidal action against *E. coli* at therapeutic doses.^[25] Fosfomycin is also a good oral treatment

option for both *E.coli* and *Enterococcus* spp. as causative agents of acute uncomplicated cystitis.^[23] Fosfomycin, which was not marketed much in India before, was notably recommended as the first line of management for UTI in guidelines given by ICMR in 2022.^[26]

Chromphenicol sensitivity was typically found to be more than anticipated. In *E.coli*, it was found to be 36% and approximately 40% sensitive to other isolates. Similar studies have reported decreasing resistance to chloramphenicol over the years.^[27] There could be several reasons for the same: its restricted use over time due to its propensity to cause adverse hematological effects, and a change in clinical practice to use it as a targeted therapy. However, the rising susceptibility does not warrant its use as a first-line drug for UTIs, mainly due to its known toxic side effects. Still, it may be considered in treating uropathogens showing multidrug resistance or serious UTIs.

Overall, a high level of resistance was observed in Fluoroquinolones and Cephalosporins (for *E.coli*, Ciprofloxacin sensitivity was 7% and the cephalosporin group approximately 10%). Various other studies have shown increasing resistance to Fluoroquinolones, which probably can be linked to their increased use as prophylactic management for uncomplicated UTIs or as empirical therapy for cases without performing culture sensitivity of the samples.^[19] Also, high resistance to Cephalosporins has been reported by multiple studies.^[17,19,20] Hence, using Ciprofloxacin for UTIs may wane off in the coming years. In a country like India, such high resistance could be attributed to several factors, such as over-the-counter use of antibiotics without proper prescriptions, non-compliance with medications, lack of implementation of antimicrobial stewardship programs, or understanding of antimicrobial patterns in a specific region. Lastly, the lack of a culture of sending microbial cultures leads to erroneous or poor prescriptions, worsening the situation even more. Overall, high sensitivity was observed for Amikacin in Enterobacteriaceae and *Acinetobacter* spp. isolates, which is consistent with the findings in a study from Kolkata (11%) and a multi-regional study of India (7.2 %).^[21,24]

CONCLUSION

The most common organism isolated in our setting was *E.coli*. Overall susceptibility pattern of uropathogen isolates showed a typical falling sensitivity in fluoroquinolones and the Cephalosporins group of antibiotics, and better susceptibility patterns for Fosfomycin, Nitrofurantoin, Carbapenems, and Aminoglycosides. The study explains the antibiogram of common isolates that might help treating physicians choose appropriate empirical treatment. The current study also highlights the importance of performing urine cultures to provide targeted therapy jointly to avoid worsening impending antimicrobial resistance. Lack of microbiological testing in many cities of India leads to no data on antibiograms or knowledge of common pathogens in those regions. This, along with irrational treatments and resistance to urinary pathogens, remains a major public health challenge for India. An effort should be made to

address this challenge in the future.

Limitations of the Study

Sample screening was not done symptom-based, which could've given a better clinical insight. Device use was not recorded, which might have helped in understanding the burden of hospital-acquired infections more clearly.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- Schaeffer AJ. Bacterial urinary tract infections in diabetes. *J Urol* 1998;160(1):293.
- Odoki M, Aliero AA, Tibyangye J, Nyabayo Maniga J, Wampande E, Kato CD, et al. Prevalence of Bacterial Urinary Tract Infections and Associated Factors among Patients Attending Hospitals in Bushenyi District, Uganda. *Int J Microbiol* 2019;2019.
- Mahdi BM, Khudhur HB, Abdul-Hussein MM. Bacterial isolates of urine and their susceptibility to antimicrobials. *Open Access Maced J Med Sci* 2020;8(A):84–8.
- Imran Sarwar M, Sarwar I, Shahbaz Hussain M, Khan Sherwani S, Hakeem A, Urooj Kazmi S. Frequency of urinary tract infection causing agents in pregnant women and their antimicrobial susceptibility profile. *J Biochem Mol Biol* 2013;46(4):107–10.
- Atia A. Antimicrobial resistance Pattern of Bacteria Isolated from Patients with Urinary Tract Infection in Tripoli city, Libya. *Asian J Pharm Heal Sci [Internet]* 2017;(December 2017). Available from: <https://ajphs.com/>
- Ahmed SS, Shariq A, Alsalloom AA, Babikir IH, Alhomoud BN. Uropathogens and their antimicrobial resistance patterns: Relationship with urinary tract infections. *Int J Health Sci (Qassim) [Internet]* 2019;13(2):48–55. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/30983946> <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC6436442>
- Flores-Mireles AL, Walker JN, Caparon M, Hultgren SJ. Urinary tract infections: Epidemiology, mechanisms of infection and treatment options. *Nat Rev Microbiol [Internet]* 2015;13(5):269–84. Available from: <http://dx.doi.org/10.1038/nrmicro3432>
- Bru JP. Infection Urinaire De L'Adulte. *Lyon Mediterr Med Med du Sud-Est* 1995;31(1):1783-1786+1789.
- Daoud N, Hamdoun M, Hannachi H, Gharsallah C, Mallekh W, Bahri O. Antimicrobial Susceptibility Patterns of Escherichia coli among Tunisian Outpatients with Community-Acquired Urinary Tract Infection (2012-2018). *Curr Urol* 2020;14(4):200–5.
- KASS EH. Pyelonephritis and Bacteriuria. *Ann Intern Med* 1962;56(1):46–53.
- K Patel DP, H. Pattani DM. A study on prevalence of bacterial isolates causing urinary tract infection at tertiary care hospital, Rajkot, Gujarat, India. *Trop J Pathol Microbiol* 2019;5(7):454–60.
- He Y, Zhao J, Wang L, Han C, Yan R, Zhu P, et al. Epidemiological trends and predictions of urinary tract infections in the global burden of disease study 2021. *Sci Rep* 2025;15(1):1–12.
- Chooramani G, Jain B, Chauhan PS. Prevalence and antimicrobial sensitivity pattern of bacteria causing urinary tract infection; study of a tertiary care hospital in North India. *Clin Epidemiol Glob Heal [Internet]* 2020;8(3):890–3. Available from: <https://doi.org/10.1016/j.cegh.2020.02.018>
- Singh VP, Mehta A. Bacteriological profile of urinary tract infections at a tertiary care hospital in Western Uttar Pradesh, India. *Int J Res Med Sci* 2017;5(5):2126.

15. Bhargava K, Nath G, Bhargava A, Kumari R, Aseri GK, Jain N. Bacterial profile and antibiotic susceptibility pattern of uropathogens causing urinary tract infection in the eastern part of Northern India. *Front Microbiol* 2022;13(August):1–9.
16. Khan N, Shahid Khan M. Prevalence of Antimicrobial Resistance in Bacterial Isolates Causing Urinary Tract Infection in Patients attending at IIMS&R Hospital, Lucknow. *Int J Life Sci Res* [Internet] 2016;2(1):1–8. Available from: <http://ijlssr.com>
17. Bhise M, Chauhan K, Anandani G, Agarwal A. Etiological Profile and Antibiotic Susceptibility of Urinary Isolates Causing Urinary Tract Infections in Patients Attending a Tertiary Care Hospital in Rajkot, Gujarat. *Cureus* 2025;17(4):2–11.
18. Moghadas AJ, Irajian G. Original Article. 2009;4(3):105–8.
19. Mandal J, Srinivas Acharya N, Buddhapriya D, Parija SC. Antibiotic resistance pattern among common bacterial uropathogens with a special reference to ciprofloxacin resistant *Escherichia coli*. *Indian J Med Res* 2012;136(5):842–9.
20. Ramesh A, Anandam S, Sateesh K, Khelgi A. Characterisation of uropathogenic *E. coli* by detecting the virulence factors and its drug resistance pattern in a tertiary care hospital in India. *Indian J Microbiol Res* 2023;10(1):33–8.
21. Mukherjee M, Basu S, Mukherjee SKM, Majumder M. Multidrug-resistance and extended spectrum beta-lactamase production in uropathogenic *E. coli* which were isolated from hospitalized patients in Kolkata, India. *J Clin Diagnostic Res* 2013;7(3):449–53.
22. Stephenson SAM, Brown P. Distribution of virulence determinants among antimicrobial-resistant and antimicrobial-susceptible *Escherichia coli* implicated in urinary tract infections. *Indian J Med Microbiol* [Internet] 2016;34(4):448–56. Available from: <https://doi.org/10.4103/0255-0857.195354>
23. Sood S, Gupta R. Antibiotic resistance pattern of community acquired uropathogens at a tertiary care hospital in Jaipur, Rajasthan. *Indian J Community Med* 2012;37(1):39–44.
24. Mohapatra S, Panigrahy R, Tak V, Shwetha J V., Sneha KC, Chaudhuri S, et al. Prevalence and resistance pattern of uropathogens from community settings of different regions: an experience from India. *Access Microbiol* 2022;4(2):1–9.
25. Dc H. No Title. (Hooper dc. Urinary tract agents: nitrofurantoin and methenamine. In: Mandell, Douglas and Bennett's principles and practice of infectious diseases. 5th edn. Vol 1. Mandell gl, Bennett je, Dolin r, editors. Philadelphia: Churchill Livingstone; 2000. P. 423).
26. Treatment Guidelines For Antimicrobial Use in Common Syndromes 2022. 2022;89. Available from: https://www.icmr.gov.in/icmrobject/custom_data/pdf/resource-guidelines/treatment_amr_2022.pdf
27. Henry Oladeinde B, Omoregie R, Olley M, Anunibe JA. Urinary tract infection in a rural community of Nigeria. *N Am J Med Sci* 2011;3(2):75–7.